
Effects of Lameness on Sow Reproduction: Nutritional Impacts

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Introduction

Lameness has long been recognized as a problem and has been associated with a decrease in sow fertility (Penny, 1980). A study conducted in Germany reported fewer litters for lame sows than nonlame sows (<3.0 litters vs. 4.5 litters; Grandjot, 2007). This author also reported higher piglet losses for lame sows than healthy sows (27% vs. 12.4%). These production losses along with early culling of young lame sows cause a financial burden to swine production estimated at \$52/sow (Grandjot, 2007). Lameness and locomotion problems are major reasons for culling (Friendship *et al.*, 1986; Jorgensen, 2000). Therefore, maintaining herd productivity through the removal and replacement of non-productive sows has proven crucial to the well-being and success of the business. The knowledge and understanding of lameness continues to grow as an increasing number of research groups work to collect and report new data on the issue. Lameness has been linked to a significant decrease in sow productivity and is known to increase the odds of early removal from the herd (Anil *et al.*, 2008). Often times, the sows that are removed for this reason are culled at a younger age than those removed for

any other reason. The early age of removal has a negative impact on litter size, piglet survival and may increase the disruption of herd health status (Dagorn and Aumaitre, 1979; D'Allaire *et al.*, 1987; Patterson *et al.*, 1997). Anil *et al.* (2009) conducted a cohort study to examine the relationship between lameness and its effects on reproductive performance and sow longevity. The results of the study showed that lame sows had litters of smaller sizes along with a smaller number of pigs born live (**Table 1**). With this, sows had a lower rate of survival 350 days after reports of lameness, and spent fewer days in the herd. Clearly, the study revealed the significant differences between lame and nonlame sows in survivability. The sow longevity also was affected by parity and farrowing performance (Anil *et al.*, 2009a). Odds ratios have been reported for sows with elongated claws, claw cracks, heel erosion and heel overgrowth. These same studies have further concluded that uneven toes significantly impact the incidence of lameness (Vestergaard *et al.*, 2006; Anil *et al.*, 2008).

Many sows have claw lesions. Surveys have shown that greater than 88% of sows have at least one claw

lesion (Anil *et al.*, 2007). These numbers can range and change due to environmental influences and aggressive behaviors depending on whether sows are housed in group pens or crates. Claw lesions may not be indicative of lameness. The lesions of greatest concern are those which penetrate the horn wall into the corium of the foot and cause an inflammatory response such as side wall cracks of the outer horn wall or white line lesions. The focus of this paper is the impact of lameness that causes pain and locomotor problems for sows. It is important for swine producers to measure and understand how and why claw lesions are occurring. This will help develop management strategies to decrease claw lesions, lameness and the risk of early removal of sows from the herd.

The objective of this paper is to build an understanding of how the metabolic and mechanistic pathways of inflammation due to lameness influence sow reproductive performance. An understanding of these mechanisms may help one to modify management practices, alter dietary formulations and implement new disease prevention strategies. These modifications should help decrease and/or prevent this possible inflammatory response as well as lead to more productive and healthier sows. This paper will mainly focus on the inflammatory response of lameness. However, the authors will use other inflammatory responses, such as starvation, mastitis and immune barrier dysfunction, to demonstrate how inflammation from lameness may cause reproductive problems.

Table 1. Effects of lameness on pigs produced per day and longevity^a.

	Not Lamé	Lamé	<i>P</i> value
Pigs born/d	0.049 ± 0.002	0.028 ± 0.003	<0.001
Survival at 350 d, %	44.6	23.6	<0.001
Total days in herd	215.7 ± 4.45	148.3 ± 10.67	<0.001

^a Adapted from Anil, *et al.*, 2009. *J. Am. Vet. Med. Assoc.* 235(6):734-738

Decreased Feed Intake and Nutrient Partitioning

Inflammation, often accompanied by pain, is one of the most apparent consequences of lameness. Many times lame sows are thin and have poor body condition. Often sows with chronic lameness have decreased feed intake. The adjustments in the sow eating habits will lead to further complications, as observed with changes in reproductive performance. Generally if a younger parity sow does not eat well, she will show a decrease in her reproductive capabilities. Younger first litter gilts are more sensitive to the negative effects of decreased feed intake during lactation than were older gilts or multiparous sows (Eissen *et al.*, 2003). Australian researchers (King and Dunkin 1986) were some of the first to demonstrate the linear relationship between daily feed intake during lactation and increased time required for sows to express estrus after weaning. Lactation is one of the most energetically expensive and challenging activities that a female can undertake. Therefore, maintaining feed intake is crucial to sow well-being and overall performance. Feed intake, especially during certain stages in the reproductive cycle, influences many of the body's systems in significant ways. For instance, the decrease of energy and protein consumption during lactation may disrupt or even change the amount of signaling of gonadotropin-releasing hormone (GnRH) from the hypothalamus. This signal impacts the

amount of LH and FSH released and subsequently affects steroidogenesis of the ovary. The reproductive effects of inadequate feed intake during lactation seem to be mediated, at least in part, through LH secretion and embryo mortality (King and Martin, 1989). Meeting sufficient levels of energy and protein through consumption of feed is crucial to the release of the hormones necessary for the proper functioning of the reproductive system.

Inflammation not only has the potential to impact feed consumption, but release of cytokines in response to inflammation will cause a change in how nutrients will be utilized and prioritized within in the body. When an inflammatory response is signaled the animal will shift where nutrients will be utilized. Processes and organs of the immune system will take priority over other production metabolism. In other words, increased energy will go to the intestinal tract, immune organs, liver, lungs and brain, while systems not as necessary for survival such as reproduction will receive decreased nutrient flow. Anabolic processes will be interrupted, and companion catabolic activities will be enhanced to help supply nutrient needs for the immune system (Spurlock, 1997). Stallmach *et al.* (1995) documented the presence of numerous cytokines in human amniotic fluid and several of the cytokines, such as IL-6, IL-8 and TNF, were found in

fetal blood. Interestingly, the presence of IL-6 and IL-8 has been associated with intrauterine growth retardation in humans. This may suggest a possible mechanism of growth retardation or compromised development in livestock species that deserves further investigation. In addition, the immune

system demands and requires a different priority of amino acids compared to those that optimize lactation, growth and reproduction. The effects of inflammation on feed intake or nutrient partitioning negatively impacts sow body composition and condition during lactation.

Body Condition

The body condition of a sow is based on a scale from 1 to 5, with 1 signifying an under conditioned body and 5 representing an over conditioned state. Often the sows or gilts with low feed intakes during lactation have a body condition score of 1 (with scores ranging from 1-5, with 1 being thin). Those pigs with lower scores often showed signs of reproductive complications. To illustrate this point, it has been seen that sows with a body condition score (BCS) of 1 have a higher frequency of acyclic ovaries than sows with a BCS of 4 (Knauer *et al.*, 2007). It is believed that for those sows with a lower BCS, changes in other body functions will likely result. A low feed intake during lactation can lead to an excessive loss of body weight, which will ultimately result in decreased sow longevity (Gaughan *et al.*, 1995) and reproductive performance (Quesnel, 2005). It is important to keep in mind the factors that contribute to a decreased BCS in lame sows. It is reasonable to account for some of the weight loss as being due to the increase in protein loss. Loss of protein causes

dramatic effects on body condition and function. Clowes *et al.* (2003) reported that a mass loss of body protein, losses greater than 9 to 12%, rapidly decreased ovarian function. Protein restriction throughout lactation alters the concentrations of circulating somatotrophic hormones and insulin at the conclusion of the lactation period. These low concentrations negatively impact post weaning ovulation rate (Mejia-Guadarrama *et al.*, 2002). It should be noted that limited follicular development and incomplete recovery of the reproductive axis at weaning seem to be the most likely causes of decreased embryonic survival in second parity sows with earlier weaning age (Willis *et al.*, 2003). For reasons similar to these, sows with inadequate feed intake during lactation increased their odds of removal from the breeding herd (Anil *et al.*, 2006). The prevention and early treatment of lameness and other claw injuries will help maintain feed consumption and appetite, leading to a decrease in reproductive complications due to lameness. It is important to note that not all sows with claw lesions will

show changes in appetite and feed consumption. The injury of the sow must be inflammatory to see many of

Cytokine Release

Severe tissue injury induces a relatively stereotypical pathophysiologic response that is manifested by catabolism, fever and other behaviors observed with illness. When an animal has an insult, most of the adjustments that happen within the body are mediated by a cascade of polypeptide molecules called inflammatory cytokines. The release of possible toxins and/or other products from cell injury drives the activation of the inflammatory cytokines. This activation leads to a variety of metabolic and endocrine changes in the body. These changes are mediated by the direct action of cytokines on tissue function and by changes in pituitary-endocrine end-organ function. Cytokines are released from immune barrier functioning cells such as endothelial

Cytokines and Reproduction

In the ovary, intrinsic cytokines IL-6, TNF α and IL-1 regulate steroidogenesis, maturation, atresia and apoptosis of ovarian cells. The release of cytokines causes a decrease in GnRH from the hypothalamus that decreases the amount of FSH and LH released from the pituitary. A severe inflammatory response from a wound may release large amounts of cytokines, such as TNF α , which will then influence

the responses described above and yet to come.

cells, monocytes, macrophages, specialized immune cells like lymphocytes and several other types of parenchymal cells. Some of the cytokines that are released are interleukins and come in the forms of IL-1, IL-2 and IL-6. Tumor necrosis factor-alpha (TNF- α), interferon-gamma (INF- γ) and several other cytokines with anti-inflammatory activity, such as IL-10 and IL-1, work in a synergistic reaction to regulate body metabolism and help the animal survive. In simplistic terms, cytokines are released to help the body in the fighting against possible toxins and other harmful byproducts of an injury. These cytokines, along with hormones, also have specific impact on the functioning of specific reproductive organs.

the ovary. The effects on the ovary will cause a reduction in steroidogenesis and even apoptosis of the ovarian cells and ultimately can result in a lost pregnancy. The most common reproductive anomaly found in cull sows was acyclic ovaries (9%, Knauer *et al.*, 2007). The occurrence of acyclic ovaries increased ($P<0.05$) as BCS of the sow decreased. Acyclic ovaries also were positively correlated ($P<0.01$) with

rear foot abscesses. There is a strong correlation between lameness and reproductive problems as just revealed with the increase of acyclic ovaries in sows with rear foot abscesses.

In relation, at the level of hypothalamus, IL-1 inhibits pulsatile secretion of GnRH, which leads to low gonadotropin secretion and low levels of sex steroids (Rivest and Rivier, 1995; Shalts *et al.*, 1991). Severe inflammatory illness induced a dramatic decrease in circulating sex steroids (Dong *et al.*, 1992) in the human male while studies with TNF α showed inhibited gonadal secretion in the mouse (van der Poll *et al.*, 1993). The interrelationships between stress, hormones and basic ovarian functions in the ovary were tested in sows (Sirotkin, 2010). This study showed that involvement of hormones (IGF-1, leptin, FSH) controls proliferation, apoptosis and secretory activity of ovarian cells. More importantly the research showed that stressors (heat stress and malnutrition) impacted proliferation, apoptosis and secretory activities of granulosa and ovarian follicular cells. Most of these responses probably are

Inflammatory States

Although the reproductive system is highlighted in this paper, one should also remember that all organ systems are altered by acute and chronic inflammatory states. There are dramatic shifts observed in liver function in livestock with an acute phase

mediated at the hypothalamic level by the induction of corticotropic releasing hormone (CRH) and/or vasopressin (VP), which together act to increase the release of the adrenocorticotrophic hormone (ACTH). The way the reproductive system responds to lack of nutrients is similar to the body's inflammatory response due to cytokine release. When the body lacks in energy, due to a decrease in feed intake, the reproductive system will often not receive the proper amount of nutrients that is needed for a high or even normal level of performance. Similarly, when cytokines are released because of an injury, the inflammatory response is activated. This system uses more and more energy to fight inflammation or insult, taking energy that would have been distributed to the reproductive system. In that way, when cytokines are released, the reproductive system has a similar response as it does to lack of energy. The distribution of energy is controlled by the brain and in essence, the metabolic response to starvation and the response to severe inflammation essentially cause similar signals within the body and impact signals from the brain similarly.

inflammatory response. Some of the changes in the liver include suppression of albumin, transferring of ceruloplasmin, and increased synthesis of proteins like fibrinogen and C-reactive protein (Dinarelo and Wolf, 1993). Many of the claw lesions and injuries

similarly fall into these types of acute and chronic inflammatory wounds and show dramatic changes in the reproductive system. The release of cytokines that accompany foot injuries ultimately leads to complications in sow reproduction; just as acute inflammatory states can lead to alterations in the liver in different species of livestock. This

Other Inflammatory Responses

Inflammation due to lameness uses certain pathways and causes certain reactions in the body that mirror other inflammatory responses such as starvation, mastitis and immune barrier dysfunction. With the release of cytokines during an inflammatory disease, a profound change in the functioning of the neuroendocrine system is observed (Reichlin, 1993; Wilder, 1995). Inflammatory cytokine-driven responses of the neuroendocrine system are similar to and resemble those reactions seen in the body during starvation; decreased thyroid function, lower levels of GH- dependent peptides and suppression of gonadal function (Reichlin, 1999).

Not only does inflammation due to lameness share commonalities with starvation, but also with the certain reproductive complications observed with mastitis. In dairy cattle, 30% of the cows with an inflammatory response to chronic subclinical mastitis showed decreased levels of circulating estradiol, timing of ovulation, follicular steroidogenesis and oocyte

link makes the impact of lameness and other foot injuries on reproduction seem much more plausible and also, leads to an understanding of why researchers report an increase in abortions, absorptions of embryos, a decrease in litter size and a lack of sows returning to estrus when sows are severely lame.

competence. These symptoms help partially explain the lower fertility of mastitic cows (Wolfenson *et al.*, 2009). The estradiol concentrations in control cows without mastitis, normal response cows with subclinical mastitis and chronic cows with subclinical mastitis showed differences that should be noted (870 ± 62 , 815 ± 127 and 269 ± 71 ng/ml, respectively) ($P < 0.01$) (Lavon *et al.*, 2009). These numbers show the significant decrease in estradiol concentrations in mastitic cows. In addition, these researchers showed that the mRNA expression for the LH receptors, cytochrome P450 and P450 7 α -hydroxylase were lower ($P < 0.05$) in chronic mastitic cows. Along with these findings, it was discovered that blastocyst formation rate was significantly lower in embryos from mastitic cows (Wolfenson *et al.*, 2009). This decrease in formation rate is similar to the inflammatory response of embryos that were obtained in laboratory from the cows during the hot summer months.

Immune barrier dysfunction is another inflammatory response that can

be studied to help in the understanding of claw inflammation. Studies showed an increase in barrier dysfunction with the impact of several factors including heat stress, water deprivation during strenuous workouts and the usage of aspirin and ibuprofen during intense exercise (Lambert *et al.*, 2002, 2007 and 2008). These studies help to understand some of the different stressors which can create inflammatory response, by increasing the amount of endotoxin that passes through the immune barrier of the intestinal tract. These different kinds of stressors can also impact keratinocyte growth and differentiation of the claw horn tissue. Release of cytokines and growth factors such as IL-1 and keratin growth factor (KGF) due to a stressor may control keratinocyte growth and differentiation through a double paracrine loop of communication between fibroblasts in the dermis and keratinocytes in the epidermis (Werner, 2001). Changes in the architecture of the horn tissue may be affected when communication between the dermal and epidermal

Impact of Nutrition

Tomlinson *et al.* (2004) wrote a valuable review on the impact of nutrition on claw health by focusing on the roles of protein, energy, macro minerals, trace minerals and vitamins on the maintenance of the claw. Trace mineral amino acid complexes from Zinpro have been shown to be more bioavailable (Wedekind *et al.*, 1992). Research also shows that trace mineral

layers is interrupted causing changes in biomechanical properties like elasticity and hardness (Hinterhofer *et al.*, 2006). Much more is known on the mechanisms of horn development in cattle than in swine at this time, but this information suggests that inflammatory responses may impact horn development in swine as well. In a small sow study, feet showing claw lesions were selected for a post mortem examination. Of the 100 feet chosen, 35 of claws were consider a lesion score 3. All feet were evaluated for the correlation between visible claw lesions and inflammation of the underlying corium, researchers found a high correlation, 90% of the lesions were associated with a pathology of the underlying corium and all of the 35 severe lesions had underlying pathology of the corium (Da Silva *et al.*, 2010a). This study demonstrates that there is a potential pathogenesis of lameness and promotes that methods of ameliorating inflammation may be useful to the swine producer.

amino acid complexes are better retained by the body (Nockels *et al.*, 1993). Supplementation of trace mineral amino acid complexes has been shown to have multiple impacts on cattle performance and health. Feeding trace mineral amino acid complexes has shown improved claw integrity (Nocek *et al.*, 2000), fertility, lactation performance (Ballantine *et al.*, 2002; Siciliano-Jones

et al., 2008), immune function expressed as somatic cell counts, (Kellogg *et al.*, 2004) and longevity (Siciliano-Jones *et al.*, 2008) in dairy cattle.

The claw health of dairy cattle when trace mineral amino acid complexes replace inorganic sources has been more variable in response at improving claw integrity (Nocek *et al.*, 2006). It has been suggested that the limited effects of trace mineral source on claw integrity may be attributed to a lower incidence of claw lesions in some studies (Nocek *et al.*, 2006) as compared with the incidence of claw

lesions observed in other studies (Ballantine *et al.* 2002; Nocek *et al.* 2000). This suggests that greater treatment responses are seen with herds with higher lesion scores. It should not be too surprising that the integrity of the claw may give us more variable responses since it has so many complex interactions with environment and metabolism. Interestingly, measurement of liver stores of the trace minerals doesn't seem to provide much of an answer either as several studies have shown the liver values are not different between inorganic treatments and trace mineral amino acid complexes (Table 2; Siciliano-Jones *et al.*, 2008).

Table 2. Effect of trace mineral source on liver mineral concentration in dairy cows, 14 wk postpartum.¹

Mineral, mg/kg of DM	Treatment		P=
	Sulfates	Complex trace minerals	
Cu	465.0 ±	492.0 ± 25	0.49
Fe	248.0 ±	227.0 ± 14	0.35
Mn	10.3 ±	10.2 ± 0.04	0.92
Mo	3.4 ±	3.4 ± 0.1	0.98
Zn	96.9 ±	105.0 ± 4.4	0.25

¹Liver biopsies were collected before trial initiation and at 14 wk postpartum. Data collected before trial initiation were used as a covariate. Least square means (±SEM) are presented.

The most important point to all this is that trace mineral amino acid complexes deliver animal performance in cattle in production and/or health. The responses may vary in magnitude depending on the specific influences of each herd, but improvement in one or more of the following parameters is

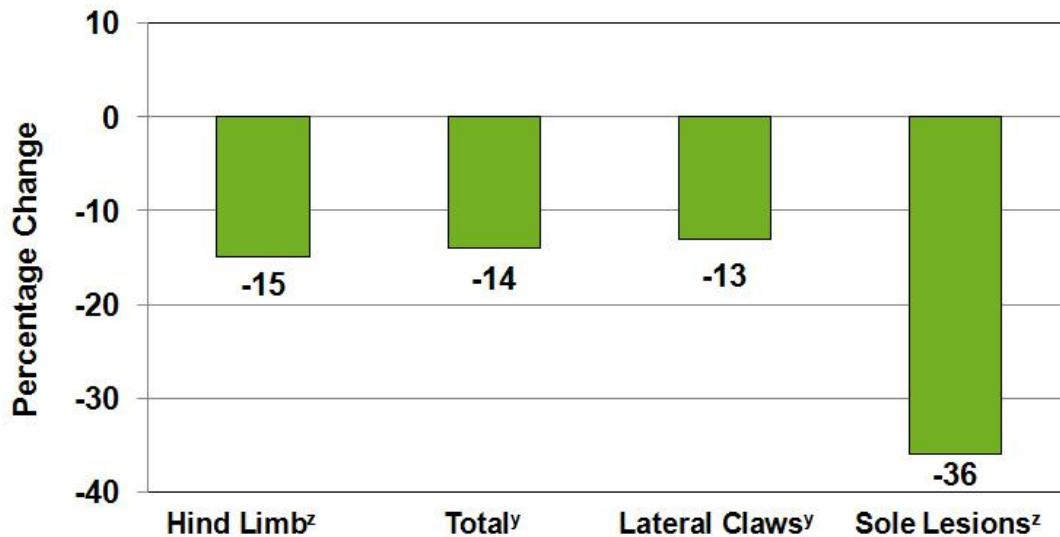
commonly reported: claw lesions, somatic cell counts, reproductive performance, milk production and longevity.

Data is not near as abundant on the issue of lameness in swine. More work has been done in the last several

years as we are starting to see many of the same responses as we see in cattle. A recent study where Zn, Mn and Cu fed as amino acid complexes (Availa[®]Sow) were supplemented to stall-housed sows in a controlled experiment, results showed a decrease in claw lesions of sows housed in gestation crates (Anil et al., 2009b). These sows were fed basal gestation and lactation diets that differed only in the source of Zn, Mn and Cu in the diet. Inorganic treatment diets supplied (Zn 125 ppm, 40 ppm Mn and 15 ppm of Cu) to the diet as sulfates. The other sows were fed diets with a partial substitution of the inorganic trace minerals with Availa-Sow (supplying 50

ppm of Zn, 20 ppm Mn, 10 ppm of Cu) with the remainder of total added levels being supplied by sulfates to make both diets iso for trace minerals added. Results showed that stall-housed sows fed Availa-Sow had fewer ($P < 0.05$) lesions after measuring lesion numbers at two consecutive gestation periods on the hind limbs and soles (**Figure 1**). These sows had fewer ($P < 0.07$) lesions on the lateral claws and total lesions. In this same study, lesion scores (severity of the lesions) were improved ($P < 0.05$) for total lesions and for total lateral claw lesions when sows were fed diets containing Availa-Sow (**Figure 2**).

Figure 1. Association of supplementing Availa[®]Sow with the number of claw lesions in stall- housed sows.

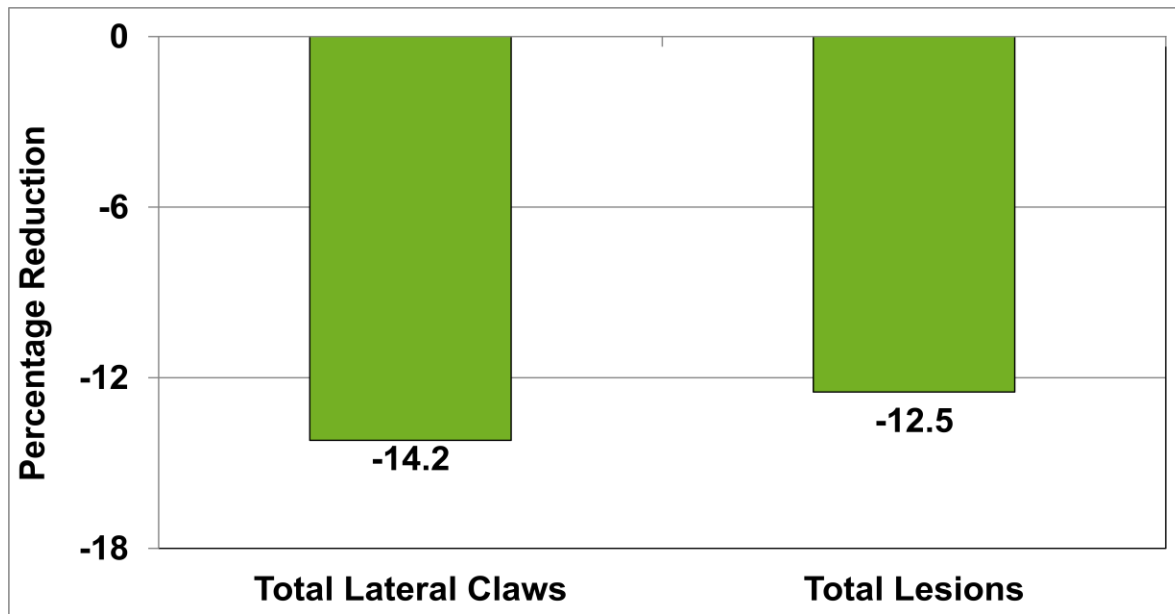


^y Trend for a decrease in lesions, $P < 0.07$
^z Decrease in claw lesions, $P < 0.05$

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SS. Anil et al., 2009. Manipulating Pig Production XII APSA. Cairns, AU p. 108

Figure 2. Improvement in severity scores when feeding Availa®Sow in stall-housed SOWS.



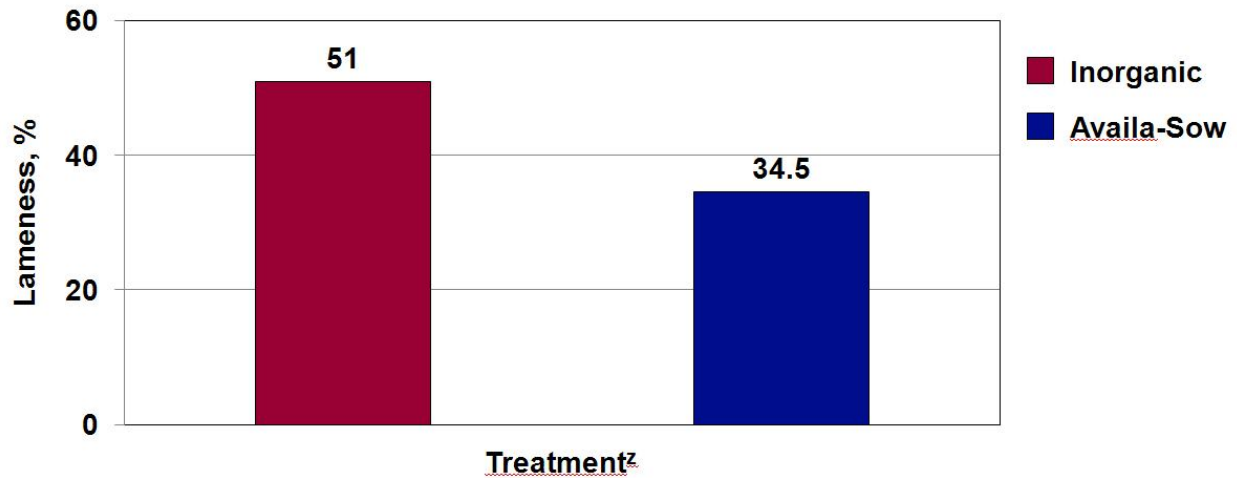
^z Decrease in claw lesions, $P < 0.05$

Anil, S. S., 2010. J. Anim. Sci. Vol. 88, E-Suppl. 88, P.p. 127, abstract. M333

Research from another project reported similar results for improvement in lesion score or severity. This study was of two different herds that had begun feeding Availa-Sow in the diets and measured the number of sows that remained the same or improved the severity of lesions with observations being taken prior to the start of ZPM treatment. A second observation was recorded in the next farrowing after feeding Availa-Sow (Da Silva, *et al.*, 2010b) and showed that all lesion

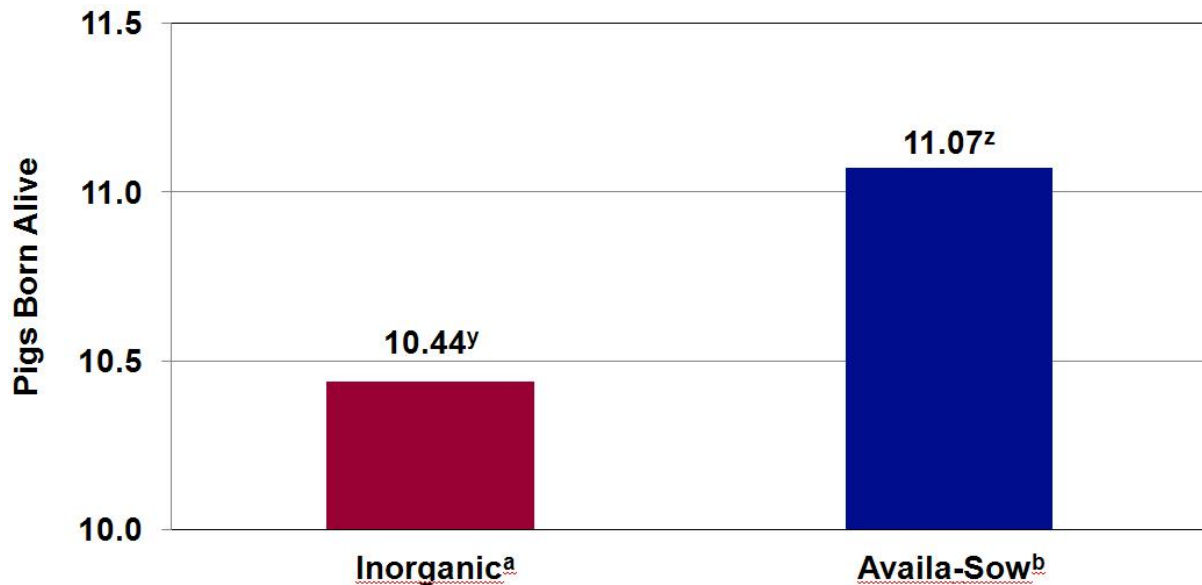
scores were better or the same ($P < 0.05$) in one farm, while in the second farm only long toes, heel sole cracks, side wall vertical cracks and side horizontal cracks were improved ($P < 0.05$). If improvements were made to inflammatory lesions then one should see changes in the number of lame sows. Analysis on prevalence of lameness was lower ($P < 0.05$) for the sows fed Availa-Sow (34% vs. 51%) over sows fed inorganic trace minerals (Anil *et al.*, 2010a)(**Figure 3**).

Figure 3. Percent lameness^a in sows when fed Availa-Sow versus inorganic trace minerals.



^a Kruskal-Wallis test
^z Reduction in lameness, $P < 0.001$
 Anil, S. S., 2010. J. Anim. Sci. Vol. 88, E-Suppl. 88, p.p. 127, abstract. M333

When reproductive performance was evaluated there were more ($P < 0.05$) pigs born alive (11.07 ± 0.21 vs. 10.44 ± 0.22 , **Figure 4**) and litter birth weight tended to be higher ($P < 0.07$) (16.99 ± 0.31 vs. 16.16 ± 0.33 , kg) (Anil *et al.*, 2010b).



^a Sulfates provided 125 ppm Zn, 40 ppm Mn, 15 ppm Cu
^b Availa-Sow; partial substitution with 50 ppm Zn as zinc amino acid complex, 20 ppm Mn as manganese amino acid complex, 10 ppm Cu as copper amino acid complex
^{y,z} Means lacking a common superscript letter differ, $P < 0.05$
 Anil, S. S., et al., 2010. Proceedings of the 21st IPVS Congress, Vancouver, Canada – July 18-21, abstract p. 862

These researchers found that group-housed sows in this experiment had higher lesions scores in all claw areas ($P < 0.05$). In the second examination of side wall cracks of group housed sows in the same experiment, results showed that the sows fed trace mineral amino acid complexes had a higher ($P < 0.05$) proportion of sows with lesions that either improved or stayed the same than the controls (91% vs. 73%) (Anil *et al.*, 2010c). These

Conclusion

Claw health is crucial to the overall well-being of the sow. If not properly treated, negative claw conditions can lead to lameness and may result in further complications. This causes a devastating loss to swine producers by decreasing reproductive performance and longevity. By improving our understanding of the factors that contribute to sow lameness and inflammation hopefully we can prevent these circumstances from occurring and avoid the many downfalls of lameness. The most important aspect of amino acid complex trace minerals is proven animal performance.

Research has shown that feeding Availa-Sow decreases the number and

researchers also showed that sows fed diets containing only inorganic sulfate trace minerals were 40% more likely ($P < 0.05$) to have ≤ 10 live born piglets compared to sows fed Availa-Sow (Anil *et al.*, 2010d). Da Silva (2010 c) showed that white line lesions and overgrown toes odds ratios of 1.151 and 1.154 respectively were positively associated ($P < 0.05$ for both lesions) with the likelihood of having ≤ 10 pigs born alive.

severity of claw lesions ultimately decreasing sow lameness. The purported mechanism of decreasing inflammatory mediators results in improved reproductive performance observed by in numbers of piglets born alive, increased litter weights, less potential for small litters, heavier weaning weights and increased feed consumption in lactation.

The economic implications for feeding Availa-Sow are a decrease in the net cost of pork production and improved profitability of the pork production enterprise.

References

- Anil, S.S., L. Anil, J. Deen, S.K. Baidoo and R.D. Walker. 2006. Association of inadequate feed intake during lactation with removal of sows from the breeding herd. *J Swine Health Prod.* 14:296-301.
- Anil, S.S., L. Anil, J. Deen, S.K. Baidoo and R.D. Walker. 2007. Factors associated with claw lesions in gestating sow. *J Swine Health Prod.* 15(2):78-83
- Anil, S.S., L. Anil and J. Deen. 2008. Analysis of periparturient risk factors affecting sow longevity in breeding herds. *Canadian J. of Animal Science:* 88(No 3):381-389.
- Anil, S.S., J. Deen, L. Anil, S.K. Baidoo, M.E. Wilson and T.L. Ward. 2009b. Evaluation of the supplementation of complexed trace minerals on the number of claw lesions in breeding sows. *Manipulating Pig Production XII, Australasian Pig Science Association.* Cairns, Australia, November 22-25, p. 108.
- Anil, S.S., J. Deen, L. Anil, S.K. Baidoo, M.E. Wilson and T.L. Ward. 2010a. Analysis of the effect of complex trace minerals on the prevalence of lameness and severity of claw lesions in stall-housed sows. *National ASAS, Denver Colorado.* *J. Anim. Sci.* Vol. 88, E- Suppl. 2, p. 827, abstract # 1014.
- Anil, S.S., J. Deen, L. Anil, S.K. Baidoo, M.E. Wilson, C. Rapp and T.L. Ward. 2010b. Comparison of the production performance of stall-housed sows receiving complexed trace minerals. *IPVS. Vancouver, Canada. Proceedings of the 21st IPVS Congress, Vancouver, Canada- July 18-21.* p.1168, abstract # P.862.
- Anil, S.S., J. Deen, L. Anil, S.K. Baidoo, M.E. Wilson, C. Rapp and T.L. Ward. 2010c. Analysis of the healing effect of complex trace minerals on claw lesions of gestating sows housed in group pens with electronic sow feeders (ESF). *IPVS. Vancouver, Canada. July 18-21.* p. 1167, abstract # P.861.
- Anil, S.S., L. Anil, J. Deen, S.K. Baidoo, M.E. Wilson and T. L. Ward. 2010d. Analysis of the association of number of piglets born alive with sow level and management factors. *J. Anim. Sci.* Vol. 88, E-Suppl. 2 P. 775, Abstract 868.
- Anil, S.S., L. Anil, J. Deen, S.K. Baidoo, M.E. Wilson and C. Rapp. 2010e. Analysis of the effect of complexed trace minerals on the prevalence of lameness and severity of claw lesions in stall-housed sows. *J. Anim. Sci.* Vol. 88, E-Suppl. 2, p. 127, abstract #M333.
- Ballantine, H.T., M.T. Socha, D.J. Tomlinson, A.B. Johnson, A.S. Shearer, and S.R. van Amstel. 2002. Effect of feeding complexed zinc , manganese, copper, and cobalt to late gestation and lactating dairy cows on claw integrity, reproduction and lactation performance. *Prof. Anim. Sci.* 18:211-218.
- Clowes, E.J., F.X. Aherne, G.R. Foxcroft and V.E. Baracos. 2003. Selective protein loss in lactation sows is associated with reduced litter growth and ovarian function. *J. Anim. Sci.* 81:753-764.
- D'Allaire, S., T.E. Stein and A.D. Leman. 1987. Culling patterns in selected Minnesota swine breeding herds. *Can. J. Vet. Res.*, 51:506.
- Da Silva, A. , J. Deen, P. Ossent and M. Wilson. 2010a. Correlation between clinically visible claw lesions in sows and inflammation of the underlying corium. *AASV. 41st Annual Meeting Proceedings. Implementing Knowledge.* March 6-9. Omaha, NE. p. 425.
- Da Silva, A., S. S. Anil, J. Deen and R. B. Morrison. 2010b. Effect of the supplementation of complex trace minerals on the healing of claw lesions in two sow

- herds. Proceedings of the 21st IPVS Congress, Vancouver, Canada. July 18-21, P. 1169, abstract #863.
- Da Silva, A., S.S. Anil, J. Deen and R. B. Morrison. 2010c. Claw lesion effects on piglets born alive of stall-housed sows in two sow herds. Proceedings of the 21st IPVS Congress, Vancouver, Canada. July 18-21, P. 293, abstract #261.
- Dagorn, J. and A. Aumaitre, 1979. Sow culling: reasons for and effect on productivity. *Livest. Prod. Sci.*, 6:167-177.
- Dinarello, C.A. and S.M. Wolff. 1993. The role of interleukin-1 in disease. *New Engl. J. Med* 328:106-113.
- Dong , Q., F. Hawker, D. McWilliam, M. Bangah, H. Bureger and D.J. Handelsman. 1992. Circulating immunoreactive inhibin and testosterone levels in men with critical illness. *Clin. Endocrinol.* 36:399-404.
- Eissen J.J., E.J. Adpeldoorn, E. Kanis, M.W.A. Verstegen and K.H. de Greef. 2003. The importance of a high feed intake during lactation of primiparous sows nursing large litters. *J. Anim. Sci.* 81:594-603.
- Friendship, R.M., M.R. Wilson, G.W. Almond, I. McMillan, R.R. Hacker, R. Pieper and S.S. Swaminathan. 1986. Sow wastage: reasons for and effect on productivity. *Can J Vet Res* 50:205-208.
- Gaughan, J.B., R.D.A. Cameron, McL. Dryden and M.J. Josey. 1995. Effect of selection for leanness on overall reproductive performance in Large White Sows. *Anim. Sci.* 60:561-564.
- Grandjot G. Claw problems cost money. *SUS-Schewinezucht und Schweinesmast. Munster-Hiltrup, Germany: Landwirtschaftsverlag GmbH*, 5:28-31.
- Hinterhofer, C., V. Apprich, J.C. Ferguson and C. Stanek. 2006. Modulus of elasticity and dry matter content of bovine claw affected by changes of chronic laminitis. *The Veterinary Journal* DOI: 10.1016/j.tvjl.2006.10.020.
- Jorgensen, B. 2000. Longevity of breeding sows in relation to leg weakness symptoms at six months of age. *Acta Vet Scand* ; 41 :105-121.
- Kellogg, D.W., D.J. Tomlinson, M.T. Socha and A.B. Johnson. 2004. Effect of feeding zinc methionine complex on milk production and somataic cell count of dairy cattle: Twelve-trial summary. *Prof. Anim. Sci.* 20:295-301.
- King, R.H. and A.C. Dunkin. 1986. The effect of nutrition on the reproductive performance of first-litter sows. 3. The response to graded increase in food intake during lactation. *Anim. Prod.* 42:119.
- King, R.H. and G.B. Martin, 1989. Relationships between protein intake during lactation, LH levels and oestrous activity in first-litter sows. *Anim. Reprod. Sci.* 19:283-292.
- Knauer, M., K.J. Stalder, L. Karriker, T.J. Baas, C. Johnson, T. Serenius, L. Layman and J.D. McKean. 2007. A descriptive survey of lesions from cull sows harvested at two Midwestern U.S. facilities. *Preventative Veterinary Medicine* 82(3-4):198-212.
- Lambert, J.C., Z. Zhou, L. Wang, Z. Song, C.J. McClain and Y.J. Kang. 2004. Preservation of Intestinal Structural Integrity by Zinc Is Independent of Metallothionein in Alcohol-Intoxicated Mice. *American Journal of Pathology*, Vol. 164, No. 6:1959-1966.
- Lambert, G.P., C.V. Gisolfi, D.J. Berg, P.L. Moseley, L.W. Oberley and K.C. Kregel. 2002. Selected contribution: Hyperthermia-induced intestinal permeability and the role of oxidative and nitrosative stress. *J. Appl. Physiol.* 92: 1750-1761.

- Lambert, G. P., M. W. Boylan, J.-P. Laventure, A. J. Bull and S. J. Lanspa. 2007. Effect of aspirin and ibuprofen on GI permeability during exercise. *Int. J. Sports Med.* 28: 722-726.
- Lambert, G.P., J. Lang, A. Bull, P.C. Pfeifer, J. Eckerson, G. Moore, S. Lanspa and J. O'Brien. 2008. Fluid restriction increases GI permeability during running. *Int. J. Sports Med.* 29:194-198.
- Lavon, Y., G. Leitner, R. Meidan, E. Klipper and D. Wolfenson. 2009. Subclinical mastitis effects on steroid concentrations and gene expression in theca cells of preovulatory follicles in cows. Joint National ASAS/ADSA meeting. Montreal, Quebec. July 11-14 *J. Dairy Sci.* Vol. 92, E-Suppl. 1
- Mejia-Guadarrama, C.A., A. Psquier, J.Y. Dourmad, A. Prunier, and H. Quesnel. 2002. Protein (lysine) restriction in primiparous lactating sows: Effects on metabolic state, somatotrophic axis and reproductive performance after weaning. *J. Anim. Sci.* 80:3286-3300.
- Nocek, J.E., A.B. Johnson and M.T. Socha. 2000. Digital characteristics in commercial dairy herds fed metal-specific amino acid complexes. *J. Dairy Sci.* 83:1553-1572.
- Nocek, J.E., M.T. Socha and D.J. Tomlinson. 2006. The effect of trace mineral fortification level and source on performance of dairy cattle. *J. Dairy Sci.* 89:2679-2693.
- Nockels, C. F., J. DeBonis, and J. Torrent. 1993. Stress induction affects copper and zinc balance in calves fed organic and inorganic copper and zinc sources. *J. Anim. Sci.* 71:2539-2545.
- Quesnel, H. 2005. Etat nutritionnel et reproduction chez la truie allaitante. *INRA Prod. Anim.* 18:277-286.
- Penny R.H.C. 1980. Locomotor dysfunction causing reproductive failure. In : Morrow, D.A. ed. *Current therapy in theriogenology*. Philadelphia : WB Saunders Co, 1980 ;1042-1045.
- Reichlin, S. 1993. Neuroendocrine-immune interactions. *N. Engl. J. Med.* 329 :1246-1253.
- Reichlin, S. 1999. Neuroendocrine Consequence of Systemic Inflammation. *Nutrition and Immune Function*. P 391-407. Washington, D.C. National Academy Press.
- Rivest S. and C. Rivier. 1995. The role of corticotropin-releasing factor and interleukin – 1 in the regulation of neurons controlling reproductive functions. *Endocr. Rev.* 16 :177-199.
- Shalts, E., Y.J. Feng and M. Ferrin. 1991. Vasopressin mediates the interleukin-1 α -induced reduced decrease in luteinizing hormone secretion in the ovariectomized rhesus monkey. *Endocrinology* 131:153-158.
- Siciliano-Jones, J.L., M.T. Socha, D.J. Tomlinson and J.M. DeFrain. 2008. Effect of Trace Mineral Source on Lactation Performance, Claw Integrity, and Fertility of Dairy Cattle . *J. Dairy Sci.* 91:1985-1995.
- Sirotkin, A.V. 2010. Effect of two types of stress (heat shock/high temperature and malnutrition/serum deprivation) on porcine ovarian cell functions and their response to hormones. *J Experimental Biology* 213:2125-2130.
- Spurlock, ME. 1997. Regulation of metabolism and growth during immune challenge: An overview of cytokine function.

- Stallmach, T., G. Hebisch, H.I. Joller-Jemelka, P. Organ, J. Schwaller and M. Engelman. 1995. Cytokine production and visualized effects in the fetomaternal unit. Quantitative and topographic data on cytokines during intrauterine disease. *Lab. Invest.* 73:384.
- Tomlinson, D.J., C.H. Mulling and T.M. Fakler. 2004. Invited Review: Formation of keratins in the bovine claw: roles of hormones, minerals, and vitamins in functional claw integrity. *J. Dairy Sci.* 87:797-809.
- van der Poll, T., J.A. Romijn, W.M. Wiersinga and H.P. Sauerwein. 1993. Effects of tumor necrosis factor on the hypothalamic-pituitary-testicular axis in healthy men. *Metabolism* 42:303-307.
- Vestergaard, K., P. Baekbo and H. Wachmann. 2006a. The effect of claw trimming on productivity and longevity of sows. *Proceedings 19th IPVS Congress, Copenhagen, Denmark.* P 482.
- Wedekind, K. J., A.E. Hortin and D.E. Baker. 1992. Methodology for assessing zinc bioavailability: Efficiency estimates for zinc-methionine, zinc sulfate and zinc oxide. *J. Anim. Sci.* 70:178-187.
- Werner, S. and H. Smola. 2001. Paracrine regulation of keratinocyte proliferation and differentiation. *Trends in Cell Biology.* 11(4):143-146.
- Wilder, R.L. 1995. Neuroendocrine-immune system interaction and autoimmunity. *Ann Rev. Immunol.* 13:307-338.
- Willis, H.J., L.J. Zak and G.R. Foxcroft. 2003. Duration of lactation, endocrine and metabolic state, and fertility of primiparous sows. *J. Anim. Sci.* 81:2088-2102.
- Wolfenson, D., Y. Lavon, R. Meidan, Z. Roth and G. Leitner. 2009. Impact of animal health on endocrinology and reproduction in dairy cows. *Tri-Annual Reproductive Symposium. Montreal, Quebec. July 11. J. Dairy Sci. Vol. 92, E-Suppl. 1*